

Original article

Constrained analogues of tocainide as potent skeletal muscle sodium channel blockers towards the development of antimyotonic agents

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Abstract

1-Benzyl-*N*-(2,6-dimethylphenyl)piperidine-3-carboxamide and 4-benzyl-*N*-(2,6-dimethylphenyl)piperazine-2-carboxamide, two conformationally restricted analogues of tocainide, were designed and synthesized as voltage-gated skeletal muscle sodium channel blockers. They showed, with respect to tocainide, a marked increase in both potency and use-dependent block.

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1. Introduction

Sodium channels are involved in many cellular functions and are altered in many pathological conditions, as in the channelopathies. Therefore, drugs targeting sodium channels constitute important therapeutic interventions for a number of diseases. Among these, tocainide (**1a**, Fig. 1), a well-known sodium channel blocker, is a class Ib antiarrhythmic drug once used in the treatment of symptomatic life-threatening ventricular arrhythmias [1,2]. It has also a marked analgesic effect in trigeminal neuralgia in humans [3,4] and antinociceptive effect in rats [5]. Furthermore, tocainide has been proposed as a clinically useful antimyotonic drug being able to block sodium channels in a use-dependent manner, i.e., with an increased potency in condition of high-frequency discharges of action potentials [6]. Myotonic syndromes are hereditary disorders of the skeletal muscle caused by missense mutations in the

human skeletal muscle sodium channel Na_v1.4 that prevent the normal fast inactivation of sodium channel [7–9]. Currently, tocainide is among the few drugs clinically used for the symptomatic treatment of muscle hyperexcitability in myotonic syndromes. However, its use as antimyotonic is hindered by unwanted adverse side-effects [10]. Thus, there is a need to develop new safer use-dependent sodium channel blockers with an improved pharmacological profile. A few years ago, a comprehensive model of the sodium channel was reported, showing that the increase in lipophilicity and molecular size of antiarrhythmic drugs can reinforce hydrophobic interactions with the binding site during use-dependent block [11,12]. As a part of our program aimed at developing new antimyotonic drugs using tocainide as a “lead compound”, a series of tocainide analogues were designed with the purpose of identifying novel potent voltage- and use-dependent skeletal muscle sodium channel blockers with very high affinity constants for the inactivated channels [13,14]. In particular, potency and use-dependent behaviour were found to be strongly increased by constraining the amino terminal group of **1a** in both a rigid alpha- and beta-proline cycle

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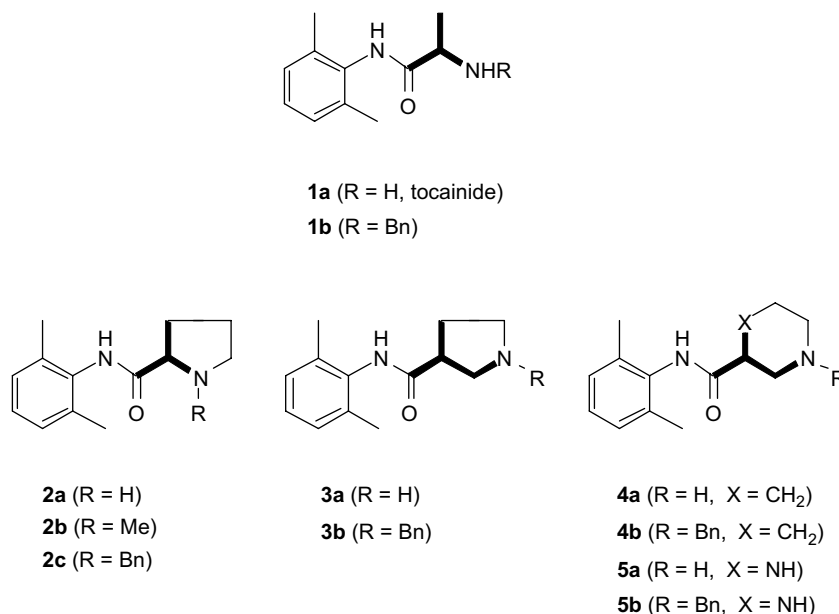


Fig. 1. Structures of tocainide analogues.

(**2a** and **3a**, Fig. 1). A further improvement was still achieved by introducing a benzyl on the amino group of proline-derived compounds (**2c** and **3b**) [15,16]. To confirm the importance of both molecular rigidity and presence of the benzyl group, we designed and synthesized two compounds, **4** and **5**, having a chemical structure that combines alpha- and beta-proline features (**2** and **3**) in a six-membered ring.

The introduction of an *N*-benzyl versus the more classic *N*-alkyl was designed as an attempt to investigate the effect of the presence of a second aromatic moiety in the molecule, which could establish specific additional hydrophobic interactions with the binding site in the protein. As we have previously reported [16], lipophilicity seems to affect the tonic block; in fact, all the *N*-alkyl derivatives showed higher tonic block values than their corresponding unsubstituted precursors. This, in turn, is related to the increase of the block potency of the channel. Recently, docking studies [17] on tocainide (**1a**) and *N*-benzyltocainide (**1b**) showed that the absence of an alkyl linked to the amino group of the tocainide led to the reduction of the van der Waals interactions with the side chain of Phe1579. In fact, the benzyl group in **1b** adopts an off-centered parallel orientation relative to the aromatic ring of the side chain of Phe1579. The additional energy of interaction explains the higher binding affinity of **1b** than tocainide (**1a**). We found that *N*-benzyltocainide analogues **2c** and **3b** showed a remarkable increase of potency, **3b** being the most active, also with respect to the corresponding *N*-methyl derivative [15]. Thus, to better clarify the role of the amino group as a pharmacophore in the drug–sodium channel interaction and confirm our previous findings, we designed two anilide derivatives, **4b** and **5b** that simultaneously resemble tocainide and alpha- and beta-proline. This has been accomplished by including the pharmacophore amino group into *N*-benzyl substituted piperidine or piperazine rings.

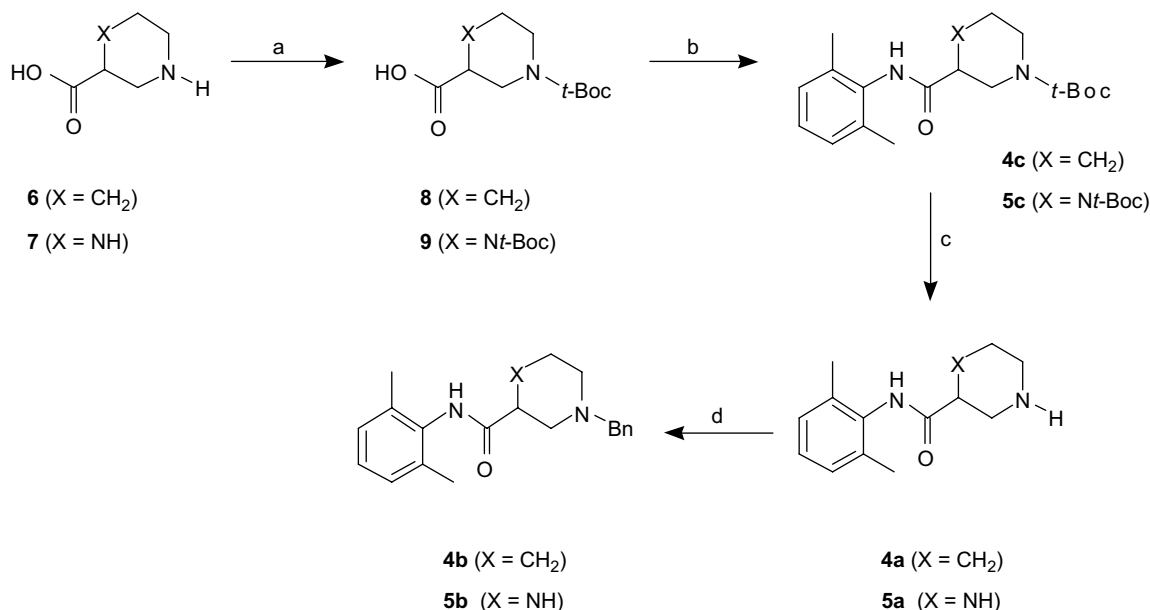
Thus, the two tocainide derived compounds were designed with the aim to merge the characteristics of **2c** and **3b**, and with the consideration that several local anaesthetics currently used in therapy, such as mepivacaine, bupivacaine and ropivacaine, are substituted 2-piperidine carboxanilides, structurally related to our compounds [18] and that 2-piperazine carboxanilides were already investigated as anaesthetic and antiarrhythmic drugs [19,20].

2. Chemistry

Compounds **4a,b** and **5a,b** were synthesized as depicted in Scheme 1. *N*-*t*-Boc derivatives **8** and **9** were prepared by reacting **6** (or **7**) and 2-*tert*-butoxycarbonylimino-2-phenylacetoneitrile (Boc-ON) in a mixture of dioxane/water in the presence of Et₃N. Then, **8** (or **9**) was reacted with 2,6-dimethylaniline in the presence of IIDQ (2-isobutoxy-1-isobutoxycarbonyl-1,2-dihydroquinoline) to afford the corresponding carboxamides **4c** and **5c**, which were, in turn, deprotected by treatment with 3 N HCl or 48% HBr to give **4a** and **5a**, respectively. The free amines were converted into their *N*-benzyl derivatives **4b** and **5b** by reacting **4a** and **5a** with benzyl bromide. We attributed the preferential benzylation of the nitrogen atom in the beta position of **5a** to several factors: the sterical hindrance of the xylylidic moiety; the electron-withdrawing effect of the same moiety that determines a reduction of basicity of the alpha nitrogen atom and, finally, the possibility of the same nitrogen atom to be involved in a five-membered pseudo-cycle with the carbonyl oxygen.

3. Pharmacology

The effects of the newly synthesized *N*-benzyl analogues of tocainide in which the asymmetric carbon atom is constrained



Scheme 1. Reagents and conditions: (a) Boc-ON, Et₃N, dioxane/water, rt; (b) 2,6-dimethylaniline, IIDQ, Et₃N, CHCl₃, reflux; (c) 48% HBr or 3 N HCl, EtOAc, rt; (d) BnBr, dioxane/water, reflux.

in a piperidine or piperazine ring were then evaluated on Na⁺ currents (*I*_{Na}) of native frog skeletal muscle fibers. Na⁺ current (*I*_{Na}) measurements executed in the presence of **4a** or **5a** were not performed because the corresponding compounds in the set of alpha- and beta-proline (**2a** and **3a**, respectively) were less active than the corresponding *N*-benzyl derivatives (**2c** and **3b**) in use-dependent block.

4. Results and discussion

The research herein described is based upon our previous works dealing with tocainide derivatives with enhanced potency and selectivity for use-dependent inhibition of the skeletal muscle Na_v1.4 channel. In particular, additional tonic potency was attained by introduction of the hydrophobic benzyl group, whereas ring constraint (proline ring) affected both potency and use-dependence of channel block. The present work shows that similar potency and use-dependence inhibition values are still possible by expanding the proline, from five- to a six-membered ring, also incorporating an additional nitrogen atom. In fact, by constraining the stereogenic center of tocainide in a rigid piperidine and piperazine cycle, as in **4b** and **5b**, respectively, a marked increase of the potency for producing both tonic and use-dependent blocks of *I*_{Na} was obtained. Tonic block IC₅₀ values of the two anilide derivatives **4b** and **5b** were similar being about 15- and 18-fold more potent than tocainide at −100 mV, respectively. During the 10 Hz stimulation, the potency of both compounds increased. Indeed, the IC₅₀ values for use-dependent block of **4b** and **5b** were about 61- and 114-fold lower, respectively (Table 1), than tocainide. In particular, a very high increase in potency for use-dependent block was observed with the **5b** analogue, with a ratio (IC₅₀ tonic block/IC₅₀ use-dependent block at 10 Hz = 31.6:2.4) of 13. In summary, the

synthesis of constrained analogues of tocainide has been proposed. The synthetic routes herein described, though not innovative and with not very high overall yields, gave access to **4b** and **5b** which present themselves as valuable pharmacological tools, both being more potent and use-dependent than tocainide. In particular, **5b** showed a marked increase in both potency and use-dependence of action; compared to tocainide, it was 18- and 114-fold more potent in tonic and phasic block experiments, respectively. In conclusion, the candidates **4b** and **5b** will be subjected to a complete pharmacological characterization as antimyotonic agents possibly overcoming tocainide's adverse side-effects. Further in vivo studies on the compounds **4b** and **5b** will be performed. Moreover, given that a stereoselective site for sodium channel blockers on adult skeletal muscle fibers has been evidenced [14], we have recently started the synthesis of the optical isomers of **4b** and **5b**; further studies are in progress to find a correlation between potency, stereoselectivity and conformational characteristics of the compounds presented in this paper.

Table 1

Concentrations for half-maximal tonic and phasic (use-dependent) blocks (IC₅₀, μM) of sodium currents, potency ratio (IC₅₀ tocainide/IC₅₀ derivatives), clog *P* and p*K*_a of tocainide (**1a**) and its analogues (**4b,5b**)

Compound	Tonic block ^a (IC ₅₀)	Phasic block ^b (IC ₅₀)	clog <i>P</i> ^c
1a (Tocainide)	580.7 ± 37.9	273.3 ± 23.3	0.76 ± 0.48
4b	38.5 ± 1.4	4.5 ± 0.3	3.53 ± 0.34
5b	31.6 ± 0.8	2.4 ± 0.4	2.81 ± 0.46

^a Tonic block: block of sodium channel at resting conditions, evaluated during infrequent depolarizing pulses.

^b Phasic block: cumulative sodium current reduction by the drug at 10 Hz stimulation frequency, obtained by concentration–response curves.

^c Calculated using Advanced Chemistry Development (ACD) Software Solaris V4.76.

5. Experimental protocols

5.1. Chemistry

Yields refer to purified products and were not optimized. All chemicals were purchased from Sigma–Aldrich or Lancaster in the highest quality commercially available. The structures of the compounds were confirmed by routine spectrometric analyses. For compounds not previously described, complete spectroscopic characterization is given; for known compounds only spectra not described in the literature are given. Melting points were determined on a Gallenkamp melting point apparatus in open glass capillary tubes and are uncorrected. ^1H NMR and ^{13}C NMR spectra were recorded on a Varian VX Mercury spectrometer, operating at 300 and 75 MHz for ^1H and ^{13}C , respectively, using CDCl_3 as solvent. Chemical shifts are reported in parts per million (ppm) relative to solvent resonance: δ 7.26 (^1H NMR) and δ 77.3 (^{13}C NMR). Absolute values of J are given in hertz. EIMS spectra were recorded on a Hewlett–Packard 6890-5973 MSD gas chromatograph/mass spectrometer at low resolution. Elemental analyses were performed with a Eurovector Euro EA 3000 analyzer. Flash chromatography was performed on silica gel (Kieselgel 60, 0.040–0.063 mm, Merck, Darmstadt, Germany) packed columns as described by Still et al. [21]. TLC analyses were performed on precoated silica gel on aluminum sheets (Kieselgel 60 F₂₅₄, Merck). PLC analyses were performed on precoated silica gel on glass plates (Kieselgel 60 F₂₅₄, Merck).

5.1.1. 1-(*tert*-Butoxycarbonyl)piperidine-3-carboxylic acid (**8**)

Yield 75%; white solid, mp: 160–162 °C, lit [22] 149–151 °C (abs EtOH/H₂O); GC–MS (70 eV): m/z (rel. int.): 229 [M^+ , 10], 57 (100); ^{13}C NMR: δ = 24.3 (1C), 27.4 (1C), 28.6 (3C), 41.3 (1C), 44.1 (1C), 45.7 (1C), 80.2 (1C), 155.0 (1C), 179.1 ppm (1C). Other spectroscopic data were in agreement with the literature [22].

5.1.2. 1,4-Bis(*tert*-butoxycarbonyl)piperazine-2-carboxylic acid (**9**)

Yield 75%; white solid, mp: 148–150 °C, lit [23] 143.0–144.5 °C for the (*S*)-enantiomer; ^{13}C NMR: δ = 28.4 (6C), 40.3 (1C), 41.6 (1C), 53.6 (1C), 54.9 (1C), 80.9 (1C), 81.3 (1C), 154.8 (1C), 155.4 (1C), 174.3 ppm (1C). Other spectroscopic data were in agreement with those reported for the (*S*)-enantiomer [23].

5.1.3. *tert*-Butyl 3-[(2,6-dimethylphenyl)amino]carbonyl]-piperidine-1-carboxylate (**4c**)

IIDQ (2.46 mL, 8.3 mmol), 2,6-dimethylaniline (0.93 mL, 7.6 mmol) and Et₃N (1.4 mL, 10.4 mmol) were successively added to a stirring solution of **8** (1.60 g, 6.9 mmol) in CHCl_3 (220 mL). The reaction mixture was heated under reflux for 6 h. The solvent was removed under reduced pressure and the residue, taken up with EtOAc, was washed three times with 2 N HCl, twice with 2 N NaOH, and then dried over anhydrous Na₂SO₄. Flash chromatography (eluent

EtOAc/petroleum ether 2:8) of the residue gave 0.39 g (17% yield) of **4c** as a white solid: mp: 59–61 °C; FT-IR (KBr): 3421, 2988, 2944, 2860, 1675, 1488, 1467, 1368, 1300, 1270, 1149, 957, 989, 858 cm^{-1} ; ^1H NMR: δ = 1.45 (s, 9H, partially overlapped to multiplet at 1.42–1.60 ppm, *t*-Bu), 1.42–1.60 (m, 1H, partially overlapped to singlet at 1.45 ppm), 1.64–1.75 (m, 1H), 1.84–1.90 (m, 1H), 2.18 (s, 6H, CH_3 -Ar), 2.40–2.60 (m, 1H), 2.95–3.15 (m, 1H), 3.25–3.40 (m, 1H), 3.65–3.85 (m, 1H), 3.95–4.10 (m, 2H), 6.95–7.10 (m, 3H, Ar), 7.35 ppm (br s, 1H: exchange with D₂O, NH); ^{13}C NMR: δ = 18.6 (2C), 24.4 (1C), 28.0 (2C), 28.6 (3C), 43.2 (1C), 44.7 (1C), 46.3 (1C), 80.3 (1C), 127.5 (1C), 128.4 (3C), 135.6 (1C), 155.2 (1C), 171.8 ppm (1C); GC–MS (70 eV) m/z (rel. int.): 332 [M^+ , 1], 121 (100).

5.1.4. Di-*tert*-butyl 2-[(2,6-dimethylphenyl)amino]-carbonyl]piperazine-1,4-dicarboxylate (**5c**)

Prepared as reported above for the synthesis of **4c**, but starting from **9**. Yield 51%; white crystals, mp: 189–190 °C (EtOAc/petroleum ether); FT-IR (KBr): 3273, 2979, 2885, 1675, 1478, 1403, 1366, 1250, 1169, 1111, 1041, 974, 868, 764 cm^{-1} ; ^1H NMR: δ = 1.44 (s, 9H, *t*-Bu), 1.50 (s, 9H, *t*-Bu), 2.19 (s, 6H, CH_3 -Ar), 2.95–3.40 (m, 3H), 3.80–4.10 (m, 2H), 4.50–4.85 (m, 2H), 6.95–7.15 (m, 3H, Ar), 7.39 ppm (br s, 1H, NH); ^{13}C NMR: δ = 18.7 (2C), 28.5 (6C), 43.3 (2C), 60.6 (2C), 80.6 (1C), 81.9 (1C), 127.7 (2C), 128.5 (3C), 135.5 (1C), 154.8 (2C), 168.0 ppm (1C); GC–MS (70 eV) m/z (rel. int.): 333 [M^+ – 100, 1], 85 (100).

5.1.5. *N*-(2,6-Dimethylphenyl)piperidine-3-carboxamide (**4a**)

To a solution of **4c** (0.40 g, 1.2 mmol) in EtOAc (9.5 mL), 3 N HCl (3.1 mL) was added. The reaction mixture was stirred at room temperature for 3 h. The solvent was removed under reduced pressure to give a white solid (0.41 g) which was recrystallized from EtOH/Et₂O to afford 0.21 g (65% yield) of the desired amine hydrochloride (**4a**·HCl): mp: 199–201 °C (abs EtOH/Et₂O), lit [18] mp: 196–210 °C (abs EtOH/Et₂O). Calcd for C₁₄H₂₀N₂O·HCl·0.17H₂O % C 61.87, H 7.91, N 10.31; found C 62.14, H 7.96, N 10.34. Compound **4a** as free amine was recovered by extraction of the corresponding hydrochloride: FT-IR (CHCl_3): 3421, 2988, 2944, 2860, 1675, 1488, 1467, 1368, 1300, 1270, 1149, 957, 989, 858 cm^{-1} ; ^1H NMR: δ = 1.51–1.62 (m, 1H, NH), 1.72–2.04 (m, 3H), 2.19 (s, 6H, CH_3 -Ar), 2.46–3.04 (m, 5H), 3.10–3.24 (m, 1H), 7.03 (s, 3H, Ar), 9.53 ppm (br s, 1H, NH-CO); ^{13}C NMR: δ = 18.8 (2C), 23.4 (1C), 27.5 (1C), 42.5 (1C), 46.6 (1C), 48.4 (1C), 127.4 (1C), 127.8 (2C), 128.6 (2C), 135.2 (1C), 173.7 ppm (1C); GC–MS (70 eV) m/z (rel. int.): 232 [M^+ , 20], 84 (100).

5.1.6. *N*-(2,6-Dimethylphenyl)piperazine-2-carboxamide (**5a**)

To a solution of **5c** (0.90 g, 2.1 mmol) in EtOAc (12 mL), 48% HBr (48 mL) was added. The reaction mixture was stirred at room temperature for 3 h. The solvent was

evaporated under reduced pressure and the aqueous phase was washed with EtOAc, made alkaline with 2 N NaOH and extracted with EtOAc. The organic phase was dried over anhydrous Na₂SO₄. The solvent was removed under pressure to give **5a** (0.22 g, 45% yield) as a white solid: mp: 172–174 °C, lit [19] 171–172 °C (CHCl₃/CCl₄); FT-IR (KBr): 3251, 3200, 3021, 2932, 2850, 2805, 1657, 1594, 1531, 1477, 1224, 1118, 897, 769, 759 cm⁻¹; ¹H NMR: δ = 1.95 (s, 2H: exchange with D₂O, NH), 2.21 (s, 6H, CH₃-Ar), 2.74–3.14 (m, 5H), 3.28 (dd, J = 3.1 and 12.1 Hz, 1H), 3.55 (dd, J = 3.1 and 8.0 Hz, 1H), 7.07 (s, 3H, Ar), 8.50 ppm (br s, 1H: exchange with D₂O, NH-CO); ¹³C NMR: δ = 18.9 (2C), 45.5 (1C), 46.5 (1C), 49.5 (1C), 59.8 (1C), 127.3 (3C), 128.4 (2C), 135.2 (1C), 170.8 ppm (1C); GC-MS (70 eV) m/z (rel. int.): 233 [M^+ , 1], 85 (100). Compound **5a** was purified by recrystallization of the corresponding hydrochloride salt, obtained by adding to the free amine a few drops of aq. HCl and then, removing the water azeotropically (toluene/abs EtOH): mp >250 °C (abs EtOH/Et₂O). Calcd for C₁₃H₁₉N₃O·2HCl·1.5H₂O % C 46.85, H 7.26, N 12.61; found C 46.84, H 7.24, N 12.68.

5.1.7. 1-Benzyl-N-(2,6-dimethylphenyl)piperidine-3-carboxamide (**4b**)

To a stirring solution of **4a** (0.15 g, 0.65 mmol) in dioxane (8 mL), a solution of K₂CO₃ (0.26 g, 1.88 mmol) in H₂O (8 mL) was added. The reaction mixture was heated to 70 °C, and then, benzyl bromide (0.09 mL, 0.76 mmol) was added dropwise. The heating was continued for 45 min. Then, the dioxane was removed under reduced pressure and the aqueous residue was taken up with EtOAc and extracted with 2 N HCl. The aqueous phase was made alkaline with 2 N NaOH and extracted twice with EtOAc. The combined organic layers were dried over anhydrous Na₂SO₄ and concentrated under vacuum to give 90 mg (43% yield) of **4b** as a white solid: mp: 133–135 °C; FT-IR (KBr): 3246, 3029, 2935, 2920, 2848, 2804, 1649, 1516, 1469, 1364, 1223, 1071, 1010, 768, 739, 699 cm⁻¹; ¹H NMR: δ = 1.55–1.80 (m, 2H), 1.90–2.25 (m, 2H, partially overlapped to a singlet at 2.15 ppm), 2.15 (s, 6H, overlapped to a multiplet at 1.90–2.25 ppm, CH₃-Ar), 2.30–2.55 (m, 2H), 2.65–2.80 (m, 1H), 2.82–3.00 (m, 1H), 3.05–3.25 (m, 1H), 3.55 (s, 2H, benzylic protons), 7.08 (s, 3H, Ar), 7.16–7.32 (m, 5H, Ar), 9.50–9.80 ppm (br s, 1H: exchange with D₂O, NH); ¹³C NMR: δ = 18.9 (2C), 22.9 (1C), 26.9 (1C), 42.1 (1C), 54.0 (1C), 55.3 (1C), 63.8 (1C), 127.0 (2C), 127.8 (2C), 128.4 (2C), 128.7 (2C), 129.6 (2C), 135.3 (1C), 137.3 (1C), 173.7 ppm (1C); GC-MS (70 eV) m/z (rel. int.): 322 [M^+ , 23], 91 (100). Compound **4b** was transformed into its hydrochloride salt by treatment with a few drops of 2 N HCl and then removing the water azeotropically (toluene/abs EtOH). The white solid so obtained (**4b**·HCl) was recrystallized from EtOH/Et₂O to afford 65 mg (67% yield) of white crystals: mp: 235–237 °C (abs EtOH/Et₂O). Calcd for C₂₁H₂₆N₂O·HCl·0.25H₂O % C 69.41, H 7.63, N 7.71; found C 69.42, H 7.68, N 7.71.

5.1.8. 4-Benzyl-N-(2,6-dimethylphenyl)piperazine-2-carboxamide (**5b**)

It was obtained as reported above for the preparation of **4b**, but starting from **5a**. Compound **5b** was obtained as a yellow oil containing also the dibenzyl derivative of **5a**, which were separated by preparative chromatography (PLC) using silica gel on glass plates (eluent EtOAc/petroleum ether 1:1) to give **5b** in 21% yield as a slightly yellowish oil: FT-IR (neat): 3356, 3201, 3066, 3031, 2983, 2942, 2822, 1676, 1601, 1497, 1445, 1373, 1303, 1248, 1145, 1047, 1028, 846, 792 cm⁻¹; ¹H NMR: δ = 1.93 (br s, 1H: exchange with D₂O, NH), 2.21 (s, 6H, CH₃-Ar), 2.31–2.44 (m, 1H), 2.50–2.66 (m, 2H), 2.88–3.02 (m, 2H), 3.04–3.16 (m, 1H), 3.54 (2d, J = 13.0 Hz, 2H, benzylic protons), 3.65 (dd, J = 7.1 and 3.6 Hz, 1H), 7.02–7.10 (m, 3H, Ar), 7.22–7.34 (m, 5H, Ar), 8.55–8.64 ppm (br s, 1H: exchange with D₂O, NH-CO); ¹³C NMR: δ = 18.9 (2C), 44.2 (1C), 53.6 (1C), 56.2 (1C), 58.9 (1C), 63.5 (1C), 127.3 (2C), 127.5 (2C), 128.4 (2C), 128.6 (2C), 129.4 (2C), 135.3 (1C), 137.7 (1C), 170.8 ppm (1C); GC-MS (70 eV) m/z (rel. int.): 223 [M^+ – 100, 1], 91 (100). Compound **5b**·HCl was obtained as reported above for **4b**·HCl in 17% yield: mp: 241–243 °C (abs EtOH/Et₂O). Calcd for C₂₀H₂₅N₃O·HCl·1.67H₂O % C 61.61, H 7.58, N 10.78; found C 61.60, H 7.55, N 10.79.

5.2. Pharmacology

5.2.1. Recording of Na⁺ current and pulse protocols

The actions of the two anilide derivatives of tocainide **4b** and **5b** were tested *in vitro* on sodium currents (I_{Na}) of single fibers of frog *semitendinosus* muscle by vaseline-gap voltage clamp method, as described in detail elsewhere [24]. The tonic block (TB) exerted by each compound was calculated as percentage reduction of the maximal peak sodium transient ($I_{Na \max}$) elicited by infrequent depolarizing steps to –20 mV from the holding potential (h.p.) of –100 mV at a frequency of 0.3 Hz. The use-dependent block exerted by each drug was evaluated by using trains of 10 ms test pulses, from the h.p. to –20 mV at 10 Hz frequency for 30 s and then normalizing the residual current at the end of the stimulation protocol with respect to the current in the absence of drug.

5.2.2. Statistical analysis

The data obtained were expressed as mean \pm standard error of the mean (SEM). The molar concentrations of each drug producing a 50% block of $I_{Na \max}$ (IC₅₀) were determined by using a non-linear least-squares fit of the concentration–response curves to the following logistic equation:

$$\text{Effect} = -100 / 1 + (K / [\text{drug}])^n$$

where effect = percentage change of I_{Na} , –100 = maximal percentage block of I_{Na} , K = IC₅₀ of tested compound, n = logistic slope factor, and [drug] = molar concentration of the tested compound [24].

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References

- [1] E.M. Vaughan, D.M. Williams, *Pharmacol. Ther.* 1 (1975) 115–138.
- [2] D.M. Roden, R.L. Woosley, *New Engl. J. Med.* 315 (1986) 41–45.
- [3] P. Lindström, U. Lindblom, *Pain* 28 (1987) 45–50.
- [4] U. Lindblom, P. Lindström, *Pain* 18 (Suppl. 1) (1984) S411.
- [5] C.J. Woolf, Z. Wiesenfeld-Hallin, *Pain* 23 (1985) 361–374.
- [6] R. Rüdel, F. Lehmann-Horn, *Physiol. Rev.* 65 (1985) 310–356.
- [7] A.L. Goldin, R.L. Barchi, J.H. Caldwell, F. Hofmann, J.R. Howe, J.C. Hunter, R.G. Kallen, G. Mandel, M.H. Meisler, Y.B. Netter, M. Noda, M.M. Tamkun, S.G. Waxman, J.N. Wood, W.A. Catterall, *Neuron* 28 (2000) 365–368.
- [8] S.C. Cannon, *Neuromuscul. Disord.* 7 (1997) 241–249.
- [9] D. Conte Camerino, D. Tricarico, J.-F. Desaphy, *Neurotherapeutics* 4 (2007) 184–198.
- [10] B. Herweg, J.S. Steinberg, *Card. Electrophysiol. Rev.* 4 (2000) 255–261.
- [11] V. Yarov-Yarovoy, J. Brown, E.M. Sharp, J.J. Clare, T. Scheuer, W.A. Catterall, *J. Biol. Chem.* 276 (2001) 20–27.
- [12] V. Yarov-Yarovoy, J.C. McPhee, D. Idsvoog, C. Pate, T. Scheuer, W.A. Catterall, *J. Biol. Chem.* 277 (2002) 35393–35401.
- [13] C. Franchini, F. Corbo, G. Lentini, G. Bruno, A. Scilimati, V. Tortorella, D. Conte Camerino, A. De Luca, *J. Med. Chem.* 43 (2000) 3792–3798.
- [14] S. Talon, A. De Luca, M. De Bellis, J.-F. Desaphy, G. Lentini, A. Scilimati, F. Corbo, C. Franchini, P. Tortorella, H. Jockusch, D. Conte Camerino, *Br. J. Pharmacol.* 134 (2001) 1523–1531.
- [15] M. Muraglia, C. Franchini, F. Corbo, A. Scilimati, M.S. Sinicropi, A. De Luca, M. De Bellis, D. Conte Camerino, V. Tortorella, *J. Heterocycl. Chem.* 44 (2007) 1099–1103.
- [16] A. De Luca, S. Talon, M. De Bellis, J.-F. Desaphy, G. Lentini, F. Corbo, A. Scilimati, C. Franchini, V. Tortorella, D. Conte Camerino, *Mol. Pharmacol.* 64 (2003) 932–945.
- [17] G.M. Lipkind, H.A. Fozzard, *Mol. Pharmacol.* 68 (2005) 1611–1622.
- [18] B. Ho, A.M. Crider, J.P. Stables, *Eur. J. Med. Chem.* 36 (2001) 265–286.
- [19] W.L. McKenzie, W.O. Foye, *J. Med. Chem.* 15 (1972) 291–295.
- [20] P.A. Tenthorey, H.J. Adams, G.H. Kronberg, B.H. Takman, *J. Med. Chem.* 24 (1981) 1059–1063.
- [21] W.C. Still, M. Kahn, A. Mitra, *J. Org. Chem.* 43 (1978) 2923–2925.
- [22] A.M. Crider, T.T. Tita, J.D. Wood, C.N. Hinko, *J. Pharm. Sci.* 71 (1982) 1214–1219.
- [23] W. Brieden, J.-P. Roduit, WO Patent 9529169, 1995. *Chem. Abstr.* 124 (1995) 176151.
- [24] A. De Luca, F. Natuzzi, J.-F. Desaphy, G. Loni, G. Lentini, C. Franchini, V. Tortorella, D. Conte Camerino, *Mol. Pharmacol.* 57 (2000) 268–277.